A population of gut epithelial enterochromaffin cells is mechanosensitive and requires Piezo2 to convert force into serotonin release


Enterochromaffin (EC) cells constitute the largest population of intestinal epithelial enteroendocrine (EE) cells. EC cells are proposed to be specialized mechanosensory cells that release serotonin in response to epithelial forces, and thereby regulate intestinal fluid secretion. However, it is unknown whether EE and EC cells are directly mechanosensitive, and if so, what the molecular mechanism of their mechanosensitivity is. Consequently, the role of EE and EC cells in gastrointestinal mechanobiology is unclear. Piezo2 mechanosensitive ion channels are important for some specialized epithelial mechanosensors, and they are expressed in mouse and human EC cells. Here, we use EC and EE cell lineage tracing in multiple mouse models to show that Piezo2 is expressed in a subset of murine EE and EC cells, and it is distributed near serotonin vesicles by superresolution microscopy. Mechanical stimulation of a subset of isolated EE cells leads to a rapid inward ionic current, which is diminished by Piezo2 knockdown and channel inhibitors. In these mechanosensitive EE cells force leads to Piezo2-dependent intracellular Ca2+ increase in isolated cells as well as in EE cells within intestinal organoids, and EE-dependent mechanosensitive serotonin release in EC cells. Conditional knockout of intestinal epithelial Piezo2 results in a significant decrease in mechanically stimulated serotonin secretion. This study shows that a subset of primary EE and EC cells is mechanosensitive, uncovers Piezo2 as their primary mechanotransducer, defines the molecular mechanism of their mechanotransduction and mechanosensitive serotonin release, and establishes the role of epithelial Piezo2 mechanosensitive ion channels in regulation of intestinal physiology.

Significance

Mechanical forces are important for normal gastrointestinal tract function. The enterochromaffin cells in the gastrointestinal epithelium have been proposed, but not previously shown, to be specialized sensors that convert forces into serotonin release, and serotonin released from these cells is important for normal gastrointestinal secretion and motility. The findings in this study show that some enterochromaffin cells are indeed mechanosensitive, and that they use mechanosensitive Piezo2 channels to generate an ionic current that is critical for the intracellular Ca2+ increase, serotonin release, and epithelial fluid secretion.


The authors declare no conflict of interest.

This article is a PNAS Direct Submission.

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This article contains supporting information online at www.pnas.org/lookup/suppl/doi:10.1073/pnas.1804938115/-/DCSupplemental.

Published online July 23, 2018.
in small and 58 ± 5% in large intestine (n = 203 ± 37 small-intestine cells per mouse, n = 74 ± 33 large-intestine cells per mouse, n = 3) (Fig. 1G). Therefore, our data suggest that Piezo2 is specific to a subset of EC cells in both small and large intestine. We used superresolution structured illumination microscopy (SIM) to examine Piezo2 subcellular distribution in EC cells (Fig. 1 H–M). We found that Piezo2 was close to cortical f-actin and in close apposition to 5-HT vesicles (Fig. 1I), and it was also in discrete intracellular vesicles that were not immunoreactive for 5-HT (Fig. 1M).

### Lineage-Traced NeuroD1 Cells Are Piezo2⁺ EE and EC Cells

EC cells constitute the largest population of EE cells (16), but recent studies suggest that the differences between EC and EE cells may be subtler than previously assumed (17, 18). Thus, we utilized NeuroD1 (19), which is a transcription factor involved in the late stages of EE cell development (18), to create a NeuroD1-cre;GCaMP5-tdTomato mouse model in which lineage-traced NeuroD1 cells expressed tdTomato (Fig. 2). We found that 80–83% of EE (CgA⁺) and 65–79% of EC (5-HT⁺) cells were NeuroD1⁺ (tdTomato⁺) in both small and large intestine, less than 4% of NeuroD1⁺ cells were CgA⁺, and we saw no NeuroD1⁺ cells in the submucosal or myenteric plexus (Fig. 2 A–C and SI Appendix, Fig. S1). Similar to the Tph1-CFP model (Fig. 1), we found that a subset of NeuroD1⁺ cells were Piezo2⁺ (Fig. 2 D–F). To examine Piezo2 expression in NeuroD1⁺ cells, we used a ribosomal trapping approach (20). We created a NeuroD1-cre;RiboTag mouse, in which a hemagglutinin (HA) tag was inserted into the coding sequence of ribosomal protein L22 of NeuroD1⁺ cells (Fig. 2 G–I) (20). We examined transcript enrichment in dissociated epithelium (Input) by qRT-PCR of HA affinity-purified or nonspecific mouse IgG control (Ms IgG). We found that HA-purified transcripts were enriched for epithelial (Vil1⁺) EE (NeuroD1⁺, CgA⁺) and EC (Tph1⁺) cell transcripts, as well as Piezo2 (Fig. 2J), but not Piezo1 (SI Appendix, Fig. S2A). These results show that NeuroD1⁺ cells are EE and EC, and their transcripts are enriched for Piezo2.

### NeuroD1⁺ Cells Have Mechanosensitive Piezo2 Currents

We used the NeuroD1-cre;GCaMP5-tdTomato (hereafter, NeuroD1-GCaMP5) mouse model to examine whether NeuroD1⁺ cells are mechanosensitive. We used a method we recently developed to establish mouse primary epithelial cultures (21, 22) and used tdTomato fluorescence to identify NeuroD1⁺ cells in primary cultures (Fig. 3A). We voltage-clamped NeuroD1⁺ cells and used a piezoelectrically driven glass force probe for mechanical stimulation by membrane displacement (Fig. 3A). We found fast mechanosensitive inward currents in 6 of 10 cells (60%) (Fig. 3B).
Nonfluorescent cells in the same preparations did not have such mechanosensitive currents (SI Appendix, Fig. S3 A and B).

The NeuroD1\(^+\) cell mechanosensitive currents were rapidly activating, reaching peak current in 3.6 ± 0.3 ms and rapidly inactivating with a time constant of inactivation \(\tau_0 = 11.4 ± 0.6\) ms at peak response (\(n = 6\)) (Fig. 3B), had nonlinear stimulus–response relationships that were fit by a two-state Boltzmann function (red) with midpoint (0.01), and for EC cell genes NeuroD1, Tph1, and NeuroD1-cre;RiboTag (magenta) with \(\mu \text{g} \text{m}^{-1} \text{m}^{-1}\) (Fig. 2). The NeuroD1\(^+\) cell mechanosensitive current biophysical properties were consistent with Piezo2 (13, 23, 24). We attempted to make a NeuroD1\(^+\) cell-specific Piezo2 knockout but NeuroD1-cre;Piezo2\(^{−/−}\) mating resulted in a lethal phenotype (SI Appendix, Fig. S4). This is likely due to involvement of NeuroD1 in neurogenesis of central nervous system neurons (25) (SI Appendix, Fig. S1).

Therefore, we used pharmacological inhibitors and Piezo2 knockdown to test whether the NeuroD1\(^+\) cell mechanosensitive currents are Piezo2. In voltage-clamped primary NeuroD1\(^+\) cells stimulated by membrane displacement, we found that mechanosensitive currents were inhibited by Gd\(^{3+}\), an established mechanosensitive ion channel blocker (23, 26) and D-GsMTx4, a Piezo1 (27) and Piezo2 (24) blocker (Fig. 3 E and F). Next we used Piezo2 siRNA to knock down Piezo2 channels. We found that compared with nontargeted (NT) siRNA, Piezo2 siRNA decreased Piezo2 mRNA by 60 ± 10\% (\(n = 4, P < 0.05\)), but not Tph1 mRNA (\(n = 4, P > 0.05\)). Piezo2 knockdown by siRNA (11, 23) abolished NeuroD1\(^+\) cell mechanosensitive currents unlike NT siRNA (Fig. 3 E and F), while both retained voltage-dependent currents (21, 28) (SI Appendix, Fig. S3C). These data show that NeuroD1\(^+\) cells have functional Piezo2 mechanosensitive ion channels.
Piezo2 Is Necessary for Mechanosensitive Intracellular Ca\(^{2+}\) Increase in NeuroD1+ Cells. We wondered whether Ca\(^{2+}\) signaling was downstream of Piezo2 activation by physiologically relevant stimuli in mechanosensitive NeuroD1+ cells. We used a rapid perfusion system to apply shear force, a highly relevant stimulus for Gl epithelium, on primary cultured NeuroD1+ cells from NeuroD1-GCaMP5 mouse (Fig. 4A), which are tdTomato+ (Fig. 4B) and GCaMP5+ (Fig. 4C). Of the stimulated NeuroD1+ cells, 58% (34 of 59) responded to 20-s shear force with a prolonged stimulus-dependent increase in intracellular Ca\(^{2+}\) of more than 200% (ΔF/F\(_{0}\)) 2.15 ± 0.5, time to peak 8.8 ± 1.2 s, n = 8, return to baseline ~60 ± 12 n = 12) (Fig. 4C and D). This was comparable to calcium stimulation by KCl (SI Appendix, Fig. S5 A–C). Shear-induced Ca\(^{2+}\) increases were reversibly inhibited by Ca\(^{2+}\)-free media, mechanosensitive channel blocker Gd\(^{3+}\), and Piezo channel inhibitor D-GsMTx4 (Fig. 4E and F). We blocked them with D-GsMTx4 or knocked them down with Piezo2 siRNA and found that as in isolated NeuroD1+ cells, shear force evoked an increase in intracellular Ca\(^{2+}\) (Fig. S5D and Movie S2). To test if Piezo2 channels were involved, we blocked them with D-GsMTx4 or knocked them down with Piezo2 siRNA and found a decrease of shear-force responses compared with controls and NT siRNA, respectively (Fig. 5E and F). These data suggest that NeuroD1+ cells are mechanosensitive within intact Gl epithelium.

Mechanically Induced EC Cell 5-HT Release Depends on Piezo2. Having established that EE cells are mechanosensitive, we wanted to know whether the 5-HT—releasing EE cells—the EC cells—are mechanosensitive, if EC cell mechanical stimulation resulted in 5-HT release, and if Piezo2 was involved. We used primary NeuroD1-GCaMP5 cultures to assay EE cell 5-HT release from NeuroD1+ cells in response to membrane displacement using a 5-HT biosensor, an engineered HEK-293 cell with genetically modified 5-HT-gated ion channel (5-HT\(_{R}\)) and GCaMP5G (Fig. 6A and B). Because 5-HT\(_{R}\) desensitizes within milliseconds, we used a genetically engineered 5-HT\(_{R}\) with large single-channel

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current and no desensitization (SI Appendix, Fig. S7 A–C) (32). We found that in 77% (17 of 22) of mechanosensitive NeuroD1+ cells from NeuroD1-GCaMP5 cultures mechanical stimulation evoked an intracellular Ca2+ increase (time to peak 2.4 ± 0.4 s, n = 5) (Fig. 6 C, E, and G, black) and 5-HT release detected by 5-HT biosensors after a diffusion-limited delay (Fig. 6 D, F, and H, black, and Movie S3). Therefore, these NeuroD1+ cells are mechanosensitive EC cells. To determine the specificity of the 5-HT biosensor responses, we used the 5-HT receptor antagonists ondansetron and found that, whereas the EC cells continued to respond to mechanical stimulus (Fig. 6 E and G, red), 5-HT biosensors did not (Fig. 6 F and H, red, and SI Appendix, Fig. S7 B and C). To test whether Piezo2 channels were responsible for mechanosensitive 5-HT release from EC cells, we used the Piezo channel blocker D-GsMTx4, Piezo2 siRNA, and NT siRNA, and found that D-GsMTx4 (green in Fig. 6) and Piezo2 siRNA (brown in Fig. 6) decreased both EC cell responses to force (Fig. 6 E and G) and the 5-HT biosensor responses (Fig. 6 F and H and SI Appendix, Fig. S7 B and C), while NT siRNA (blue in Fig. 6 E–H) did not. Piezo2 siRNA-treated EC cells retained their chemosensitive responses to TRPA1 agonist AITC (SI Appendix, Fig. S7 D and E). These results suggest that in mechanosensitive EC cells, force-induced Piezo2-dependent intracellular Ca2+ increase leads to 5-HT release.

**Epithelial Piezo2 Knockout Decreases Mechanosensitive Epithelial Secretion.** To test the EC cell Piezo2 role in GI physiology, we used the GI epithelial driver Vlcre (SI Appendix, Fig. S8) to create conditional GI epithelium-specific Piezo2 knockout Piezo2 CKO (VlcrePiezo2 CKO) (SI Appendix, Fig. S9) and compared it to littermate control Piezo2 WT (Piezo2 WT). To determine whether epithelial Piezo2 contributes to pressure-induced secretion, we used a custom Ussing chamber, in which we are able to apply simultaneous pressure- and voltage-clamps (11) and measure pressure-induced epithelial short-circuit currents (I sc), which is a surrogate for epithelial secretion (Fig. 7A) (33). We tested the hypothesis that luminal mechanical stimulation leads to EC cell Piezo2-dependent epithelial fluid secretion by a previously established secretion circuit that relies on epithelial 5-HT (Fig. 7B) (3, 4, 11). When we pressurized the epithelial side in steps of increasing amplitude in Piezo2 WT, we found that increasing pressure resulted in a stimulus-dependent I sc increase (black traces in Fig. 7 C and D). Compared with Piezo2 WT, the Piezo2 CKO mice had a significantly decreased pressure-induced I sc increase (black traces in Fig. 7 C and D). Secretion response to acetylcholine was not different between Piezo2 WT and Piezo2 CKO (n = 15, P > 0.05) (Fig. 7C). Inhibition of pressure-induced secretion was not complete in Piezo2 CKO compared with Piezo2 WT. Thus, we compared the pressure-induced secretion responses to inhibitor D-GsMTx4 in both models. We found that D-GsMTx4 inhibited pressure-induced secretion response in Piezo2 WT (P2 WT) but not in Piezo2 CKO (P2 CKO) (Fig. 7 E and F). These results suggest that EC cell Piezo2 is important for mechanically induced epithelial secretion.

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**Fig. 5.** NeuroD1+ cells within intact epithelium depend on Piezo2 for mechanosensitivity. (A) Confocal imaging of a typical planar organoid generated from NeuroD1-cre;GCaMP5-tdTomato mouse small intestine with an EC cell (magenta), phalloidin (cyan), and DAPI (blue) labeling. Monolayer organization is demonstrated by orthogonal views derived from the areas highlighted by the dashed cross-hairs to the right (vertical line) and below (horizontal line). (B) Piezo2 immunofluorescence (yellow), with C, colocalization of Piezo2 and tdTomato labeling. Voxel size x,y = 265 nm; z = 350 nm. [Scale bar (applies to A–C), 5 μm.] (D) Overlaid DIC/epifluorescence image of planar epithelial organoids showing EC cells (tdTomato, Right) and GCaMP5 at rest (Top), during shear force stimulation (Middle), and after recovery (Bottom). (E) Representative Ca2+ (GCaMP5) responses (ΔF/Fo) to shear-force stimulation of NeuroD1+ cells within planar organoids with Control solution (black), in the presence of 10 μM D-GsMTx4 (green) and transfected with Piezo2 siRNA (brown) and NT siRNA (blue). (F) Individual (circles) and mean ± SEM (bars) peak Ca2+ (ΔF/Fo) responses for Control shear (2.7 ± 0.5, n = 8, black), 10 μM D-GsMTx4 (0.3 ± 0.05, n = 4, green), Piezo2 siRNA (P2 siRNA, 0.06 ± 0.02, n = 8, brown), and NT siRNA (2.9 ± 0.5, n = 5, blue) in planar organoids (*P < 0.05 paired t test for Gd3+ and D-GsMTx4 vs. Control, and unpaired t test for Piezo2 siRNA vs. NT siRNA).
Discussion
Orchestrated GI secretion and motility are critical for normal digestion. These GI functions are regulated by the coordinated responses to intraluminal nutrients and mechanical forces. Epithelial EE and EC cells sense nutrients and metabolites, and coordinate physiologic responses (28, 34). EC cells, the most common type of EE cells (16), were proposed to be specialized mechanosensors that, in response to force release 5-HT (1, 2), which stimulates secretory reflexes (3, 4). However, EC cell mechanosensitivity was inferred from studies on intact tissues (2, 35) and cell models (9–11, 24). In this study we addressed critical knowledge gaps: whether EE and EC cells are mechanosensitive and, if so, what is the mechanism of their mechanosensitivity?

We found that mechanosensitive ion channel Piezo2 was specific (~80%) for EC cells in both small and large intestine of a Tph1-CFP mouse model (15), which is similar to Piezo2 specificity for human small-intestine EC cells (11). Interestingly, Piezo2 was not present in all EC cells: Piezo2+ EC cells constituted a subset of ~60–70% of EC cells. Recent studies show that the differences between EC and EE cells may be subtler than previously assumed (18, 36). EC cells express enzymes required to produce signaling molecules other than 5-HT, such as secretin and substance P (18, 36), and even contain endocrine hormones, such as CCK, GLP-1, and somatostatin, that were previously ascribed to other EE cells (37). Thus, we lineage-traced all EE cells using NeuroD1Cre; because NeuroD1 is a transcription factor that is a “late” determinant for EE cell differentiation (18, 38). We found that NeuroD1+ were EE and EC cells and, importantly, they did not include non-EE secretory cells, such as goblet and Paneth cells. Like in the Tph1-CFP mouse, we found that a subset of NeuroD1+ EC cells was Piezo2+, and that Piezo2 mRNA, as well as EE and EC cell-specific mRNAs, were enriched within NeuroD1+ cell transcripts. These results suggested that a well-defined subset of Piezo2+ EC and EE cells may be mechanosensitive. Consistent with the immunofluorescence, lineage tracing, and expression results, functional experiments showed that ~60% of primary EE cells had Piezo2-dependent mechanosensitivity when examined by electrophysiology and Ca2+ imaging. Furthermore, using 5-HT biosensors and the EC cell-specific TRPA1 chemical activator (28, 29), we found that mechanosensitive Piezo2+ 5-HT-releasing EC cells made up the largest subset (~80%) of mechanosensitive EE cells.

EC cell 5-HT has neuroepithelial (28) and endocrine (7, 39) roles. Mechanosensitive 5-HT release from intestinal mucosa has a rapid-rise phase measured in milliseconds (2, 40), presumably for neuroepithelial communication (28). This speed is consistent with the fast Piezo2 kinetics (23, 41). We found that EC cell Piezo2 was distributed at high densities near cortical f-actin, suggesting membrane localization, and close to 5-HT vesicles, which suggested functional coupling. Mechanosensitive EE cells had nonrectifying mechanosensitive currents that activated and inactivated within tens of milliseconds. These mechanosensitive currents resembled heterologously expressed Piezo2 currents (23), and Piezo2 currents in Merkel cells (13, 42) and neurons (43, 44). They were inhibited by Piezo pharmacological blockers and Piezo2 knockdown by siRNA. Furthermore, Piezo2-dependent mechanosensitive 5-HT release by EC cells was also fast, limited only by diffusion.

In addition to the rapid onset, EC cell 5-HT release also has a prolonged phase lasting many seconds (2, 40), which likely contributes to EC cell endocrine function (7, 39). The lengthy 5-HT release in response to force was surprising given rapid inactivation of Piezo2 (23, 41). Because the previous studies were done in intact tissues, it was possible that the prolonged responses to force depended on the enteric nervous system (ENS) (40, 45). However, we found that mechanical stimulation by rapid membrane displacement (50 ms) or longer-lasting shear stress (20 s) of isolated EE cells in intact intestinal organoids that lack ENS, resulted in intracellular Ca2+ increases that lasted many seconds and depended on Piezo2 and extracellular Ca2+. When we focused on EC cells using biosensors to detect single-cell 5-HT release (46), we again found that brief EC cell mechanical stimulation led to Piezo2-dependent intracellular Ca2+ increase and was followed by 5-HT release that was rapid in onset and lasted several seconds. These findings suggest that mechanosensitive EC cells amplify Piezo2 responses to force into temporally controlled 5-HT release.

Fig. 6. Mechanosensitive EC cell 5-HT release depends on Piezo2. (A) 5-HT biosensor experiment showing overlaid DIC/tdTomato images with an EC cell, and (B) GCaMP5 in both the EC and 5-HT biosensor cell (lower part of image). EC cell mechanical stimulation by force probe results in (C) Ca2+ increase in EC cell, and (D) later in 5-HT biosensor. (E) Representative traces of Ca2+ responses (ΔF/F0) during EC cell mechanical stimulation, and F, resulting 5-HT biosensor activity in a control experiment (black), with 0.1 μM ondansetron (red), 10 μM D-GsMTx4 (green), Piezo2 siRNA (brown), and NT siRNA (blue). Vertical lines represent stimulation of EC cell (dashed) and initiation of 5-HT biosensor cell response (dotted). (G) Individual (NeuroD1+ cells triangles, 5-HT biosensors circles) and mean ± SEM (bars) Ca2+ responses (ΔF/F0) of EC cell mechanical stimulation and (H) resulting 5-HT biosensor cell activity in controls (EC cell: 3.2 ± 0.5, n = 22 and 5-HT biosensor: 2.7 ± 0.6, n = 22), with ondansetron (EC cell: 3.9 ± 0.5, n = 5 and 5-HT biosensor: 0.5 ± 0.1, n = 5), with D-GsMTx4 (EC cell: 0.1 ± 0.03, n = 6, and 5-HT biosensor: 0.19 ± 0.09, n = 6), with Piezo2 siRNA (EC cell: 0.6 ± 0.2, n = 13, and 5-HT biosensor: 0.7 ± 0.2, n = 13), and with NT siRNA (EC cell: 2.9 ± 0.4, n = 15, and 5-HT biosensor: 3.3 ± 0.7, n = 15) (P < 0.05 when comparing D-GsMTx4 to control, and Piezo2 siRNA to NT siRNA for EC cell and biosensor cell, and when comparing ondansetron and D-GsMTx4 to control for biosensor cell only, by unpaired t test).
The mechanotransduction mechanism linking Piezo2 with intracellular Ca\(^{2+}\) increase and 5-HT release requires further clarification. Because EE and EC cells are electrically excitable (21, 28, 47), it is possible that Piezo2 generates a receptor potential that activates voltage-gated sodium channels, which mediate seconds-long EC-cell-bursting activity (21, 28), and activate voltage-gated calcium channels, which have been implicated in EC cell chemosensitivity (28, 48, 49). Another possibility is that Ca\(^{2+}\) flux through Piezo2 directly activates EC cell Ca\(^{2+}\)-induced Ca\(^{2+}\) release (50) or ATP release and subsequent autocrine P2X activation (51). These possibilities are not mutually exclusive, and clarifying the involvement of these mechanisms in EC mechanotransduction may help delineate EC cell roles within the neuroepithelial (28, 52) and endothelial (7, 39) systems.

To examine the physiological role of Piezo2 in EC cell mechanosensitivity, we made a conditional GI epithelium Piezo2 knockout (Piezo2\(^{CKO}\)). EC cell 5-HT is important in pressure-induced secretion responses (3, 4), and maximal pressure increases both 5-HT release and short-circuit current (\(I_{sc}\)), a surrogate for epithelial secretion (11). Thus, we compared with Piezo2\(^{WT}\), the Piezo2\(^{CKO}\) pressure- and voltage-clamped small intestine, and found that Piezo2\(^{CKO}\) had a diminished pressure-induced short-circuit current at multiple pressures. Pressure sensitivity was not further decreased by Piezo blocker D-GsMTx4 in Piezo2\(^{CKO}\) compared to Piezo2\(^{WT}\). Our results suggest that EC cell Piezo2 is responsible for a portion, but not all, pressure-induced secretion response. This is intriguingly akin to the results found in somatosensory light touch, where Piezo2 deletion in both Merkel cells and afferent neurons is required for complete sensory loss (43). In the GI system, EC cells communicate with intrinsic primary afferent sensory neurons (IPANs), which were previously described to be mechanosensitive (53). Further work is required to determine the identity of the IPAN mechanotransducer, and whether inhibition of IPAN and EC cell mechanosensitivity leads to a more complete mechanosensory loss.

In summary, we show here that subsets of primary EE and EC cells are mechanosensitive, that their mechanosensation requires Piezo2 channels, the activation of which by force leads to a rapid ionic current, mechanosensitive intracellular Ca\(^{2+}\) increase, mechanosensitive 5-HT release from EC cells, and pressure-induced epithelial fluid secretion.

**Methods**

All experimental procedures were approved by the Institutional Animal Care and Use Committee of the Mayo Clinic.

**Drugs.** Gadolinium (Gd\(^{3+}\)), 5-HT, AITC, and ondansetron (Sigma-Aldrich), Yoda1 (21904; Cayman Chemical) were all made as stock solutions (1 mM) in water. Working solutions were prepared from stock on the day of the experiments, and D-GsMTx4 (provided by Philip Gottlieb, State University of New York, Buffalo, NY) was made as a working solution on the day of the experiments.

**Animals.** Piezo2\(^{WT}\) mice were provided by Ardem Patapoutian, The Scripps Research Institute, San Diego, CA. NeuroD1::cre and Thp1::CFP mice were provided by Andrew Leiter, University of Massachusetts, Worcester, MA. Mouse lines were obtained from The Jackson Laboratories: WT and Piezo2\(^{CKO}\) conditional GI epithelium and Piezo2\(^{WT}\), (Invitrogen), 1% L-glutamine (Invitrogen) in dishes (MatTek Corporation).

**Cultures.** Primary murine colon dissociation. Primary murine colon dissociation was similar to recently described work (21, 22); NeuroD1::GCaMP5 mice were killed at 5-7 wk and a 10-cm length of colon was removed. Full-thickness tissue was inverted, chopped, and washed three times in ice-cold PBS. The tissue was digested under agitation at 37 °C in DMEM (Sigma), 0.1% BSA (Sigma), and 0.6 mg/mL Collagenase type XI (C9407; Sigma) in four separate digestions, for a total of 46 min. Supernatants were collected from the last two digestions, spun twice at 100 g for 5 min, and suspended at 1,000,000 cells per milliliter in DMEM, 5% heat-inactivated FBS (F4135; Sigma), 1% Pen-streptomycin, 1% Glutamine (Invitrogen) in dishes (MatTek Corporation) coated with 5% (w/vol) Matrigel (Corning). Cells were maintained in standard culture conditions for 24–48 h. siRNA transfection efficiency was optimized using siGLO Green Transfection Indicator (D-001630-01-05; Thermo Scientific) transfected using Lipofectamine 2000 (Invitrogen). For primary cell culture siRNA experiments, FBS and Matrigel concentrations were dropped to 2.5%. When noted, 20 nM siRNA Accell Mouse Piezo2 siRNASMARTpool (E-163012-00-0005; Thermo Scientific) or 20 nM Accell Nontargeting Control siRNA (D-001910-10-05; Thermo Scientific) were transfected for 48–72 h.

**Marine Jejunum Organoids.** Organoids were cultured and maintained according to Intesctult Organoid Growth Medium (StemCell) instructions. NeuroD1::GCaMP5 mice were provided by Andrew Leiter, University of Massachusetts, Worcester, MA. Mouse lines were obtained from The Jackson Laboratories: WT and Piezo2\(^{CKO}\) conditional GI epithelium and Piezo2\(^{WT}\), (Invitrogen), 1% L-glutamine (Invitrogen) in dishes (MatTek Corporation).
mice were killed at 8–10 wk, and a 20-cm length of small intestine was removed. The segment was cut on glass. washed, flushed with PBS, and then cut into 2-cm pieces. The tissue pieces were resuspended in PBS and pipetted with fresh buffer until the supernatant was clear. Tissue pieces were resuspended in 1 ml of Cell Dissociation Buffer (STEMCell) and incubated on a rocking shaker at room temperature for 15 min. Dissociation reagent was removed and PBS containing 0.1% BSA was added to the tissue. Tissue pieces were then pipetted up and down three times in the buffer and this was repeated four times to generate different four fractions. The first two fractions were discarded and the last two fractions were collected, and spun at 300 x g. Both fractions were combined in DMEM/F12 (Sigma), counted, spun again, and resuspended in equal parts of room temperature Intesticult Organoid Growth Medium and Matrigel. Tissue was shaken vigorously, spun at 1,000 x g. Tissue fragments were then resuspended in prewarmed 24-well dishes (Corning).

Organoids were grown under standard cell culture conditions and Intesticult Organoid Growth Medium was replaced every 3 d. After 7 d in culture, the organoids were passaged. Gentle Cell Dissociation Reagent was used to break up the organoid domes and then incubated on a rocking shaker at room temperature for 10 min. The organoid fragments were spun at 300 x g, resuspended in DMEM/F12, counted, spun again, and resuspended in equal parts of room temperature Intesticult Organoid Growth Medium and Matrigel. Tissue was shaken vigorously, spun at 1,000 x g. Tissue fragments were then resuspended in prewarmed 24-well dishes (Corning).

Gene Expression.
RNA isolation. Ribotag protocols were modified from previous studies (20). Colon tissue was harvested from 6- to 10-wk-old Neur01-RiboTag mice. Epithelial tissue was isolated by placing full-thickness tissue in Dulbecco’s PBS supplemented with EDTA (2 mM) and DTT (1 mM) and incubated at 37 °C for 20 min. Tissue was shaken vigorously, spun at 1,000 x g. Tissue fragments were then resuspended in prewarmed 24-well dishes (Corning).

The extracellular solution contained: 150 mM Na+, 2.5 mM Ca++, 1 mM Mg2+, 160 mM Cl−, 5 mM Hepes, and 5.5 mM glucose, pH 7.3, 300 mmol/kg; the intracellular solution contained: 140 mM Cs+, 150 mM Cl−, 4 mM Mg2+, 2 mM Ca++, 10 mM Hepes, and 5 mM EGTA, pH 7.3, 300 mmol/kg.

Calcium Imaging. Primary cultures from NeuroD1-Cre;GCaMP5-MdTomato mice were dissociated and grown for 24 h in MatTek dishes (MatTek Corporation) as above. Bath solution contained: 150 mM NaCl, 5 mM KCl, 2 mM MgCl2, 2 mM CaCl2, 10 mM Hepes, and 10 mM Hepes, pH 7.3, 320 mmol/kg (adjusted with sucrose) and viscosity 0.05 by Student t tests, as specified in the text, and specifically two-tailed unpaired t tests with Welch’s correction (Control vs. treatment) (Fig. 3) using GraphPad Prism v6 (GraphPad Software, https://www.graphpad.com).
unless otherwise indicated. Blockers were preapplied through the bath and coapplied during the shear. Ca^{2+}‐free bath solution, GD_{30} (30 μM), D‐GluMTx4 (10 μM), and AITC (150 μM) solutions were freshly prepared on the day of the experiments. All experiments were performed at room temperature (25 °C). Fluorescence time series were converted to 7'AM Assay (ΔF/F_0 = (F – F_0)/F_0), where F_0 is the baseline fluorescence for each trial.

5-HT Biosensor Experiments. High‐conductance nondenaturing 5‐HT IR was a kind gift of Cecilia Bouzat, Instituto de Investigaciones Bioquímicas de Bahía Blanca, Argentina (32). HEK293 cells were cotransfected with 5‐HT IR and GCaMP5G (1 μM; Addgene #31788) and cocultured with primary EC cells for 24 h before the experiments. Primary EC cells were mechanically stimulated using a fire‐polished glass micropipette (3‐μm indentation, 50‐ms duration) driven by a piezotransducer P‐621‐TCD attached to an E‐625 CR controller (Physik Instrumente). D‐GsMTx4 (10 μM), 5‐HT (1–10 μM), and ondansetron (0.1 μM), Yoda1 (0 μM), and AITC (150 μM) were freshly made on the day of the experiments. Acquisition and experimental conditions were as described in the previous section.

Using Chamber. Solutions. The Krehs‐Ringer solution consisted of: 120 mM NaCl, 5.9 mM KCl, 15 mM NaHCO3, 1.6 mM NaH2PO4, 1.3 mM CaCl2, 2.4 mM MgCl2, pH 7.4, gassed with 95/5 mixture of O2/CO2 (25 °C). Fluorescence time series were converted to the baseline fluorescence for each trial.

Tissue preparation. Segments of jejunum (4 cm) were cut along the mesenteric border, and luminal contents were gently removed. Tissue was cut into 2‐cm segments. During preparation, the tissues were bathed in ice‐cold Krehs‐Ringer solution.


Short‐circuit current measurements with mechanical pressure. The full‐thickness preparations of mouse jejunum with a cross‐sectional area of 0.3 cm2 were mounted in 4 mL Using chambers (Physiologic Instruments). Transepithelial potential difference was measured using paired Ag‐AgCl electrodes via 3% agar with 3‐M KCl bridges and clamped at 0 mV by another pair of Ag‐AgCl electrodes. The mucosal and serosal surfaces of the tissue were bathed with 4 mL of Krebs‐Ringer solution with mannitol and glucose, respectively, maintained at 37 °C during the experiments. Tissue equilibrated to attain stable basal short‐circuit current (Isc) and tissue conductance (Gt) for 30 min before conducting the experiment. Hydrostatic pressure was applied using DAPK pneumatic transducer (Bio‐Tek Instruments) in a sealed mucosal chamber. Pressure stimuli of 10 s duration were applied from rest (atmospheric pressure). To assure tissue viability, acetycholine (100 μM) was applied to the serosal side at the end of the experiment. Tissue was not used if there was no response to acetycholine. Data were recorded using Acquire and Analyze 2.3 (Physiological Instruments). Data analysis. Raw data were exported into text format and uploaded into Clampfit 10.5 (Molecular Devices). Pressure‐induced peak short‐circuit current (Isc) was measured by subtracting the baseline fluorescence for each trial.

ACKNOWLEDGMENTS. We thank Mrs. Lyndsay Busby for administrative assistance, and Mr. Eugene Kruger and the Mayo Clinic Optical Core for assistance with confocal imaging of organs. This work was supported by Grant NIH DK106456, Pilot and Feasibility Grant from the Mayo Clinic Center for Cell Signaling in Gastroenterology (NIH DK084567), and a 2015 American Gastroenterological Association Research Scholar Award (AGA RSA) (all to A.B.); NIH Grants DK100223 and DK110614 (to A.B.L.); and NIH Grant DK057266 (to G.F.).